



Advanced simulation protocols in GROMACS

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Ensemble scaling in biomolecular simulations

transitions in biomolecules are (very) infrequent

but transitions are often fast



can (smartly) combine sampling for many shorter simulations

Different problems, trivial parallelism

- Different drug molecules binding to the same protein
- Different mutations to the same protein

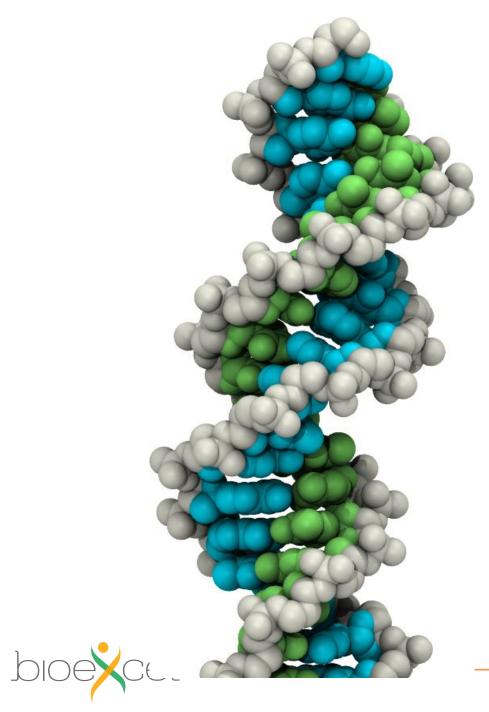
Ensemble parallelism

Processes in cells and experiments involve ensembles of large numbers of molecules

We can use ensembles in simulations:

- Do N simulations instead of 1 (trivial, common practice)
- Combine information from N simulations to get answers faster
 - Markov state modelling
 - Aggregating data from multiple walkers

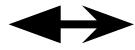




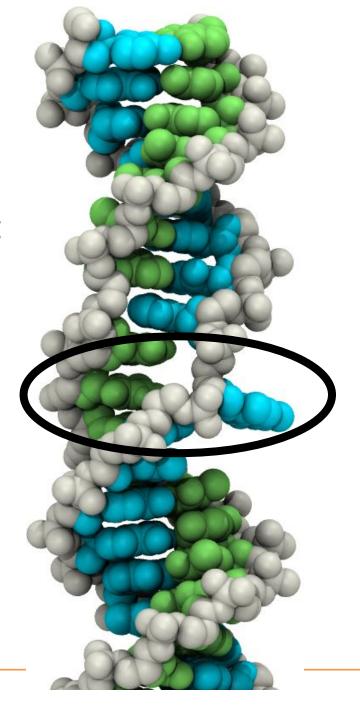
Example study:

DNA base pair opening

1 ms⁻¹



closing 10⁴ ms⁻¹



Different Approaches

General exploration

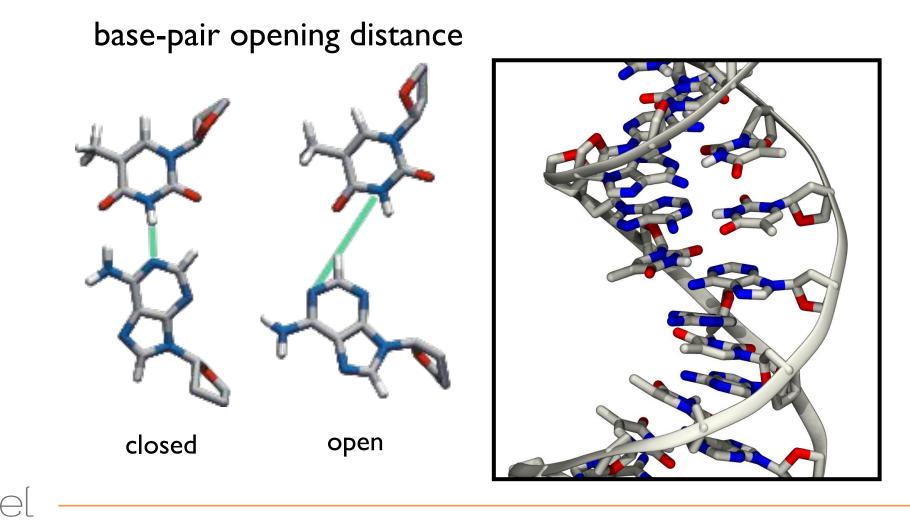
- Simple MD simulation
- Replica exchange protocols
- "Flooding"
- Conformational sampling

When you know your reaction coordinate

- Pulling
- Accelerated weighted histogram method

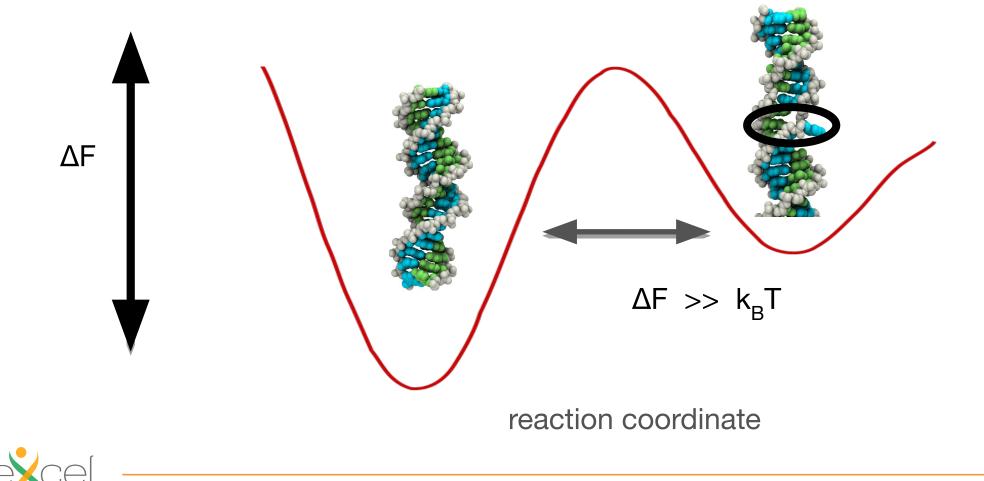


A reaction coordinate to enhance sampling

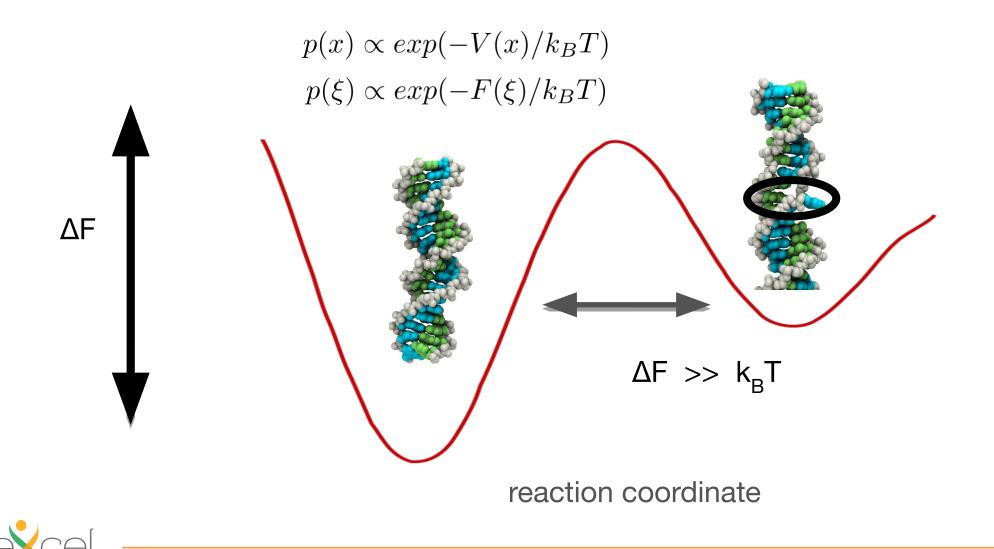


Sampling a Boltzmann distribution:

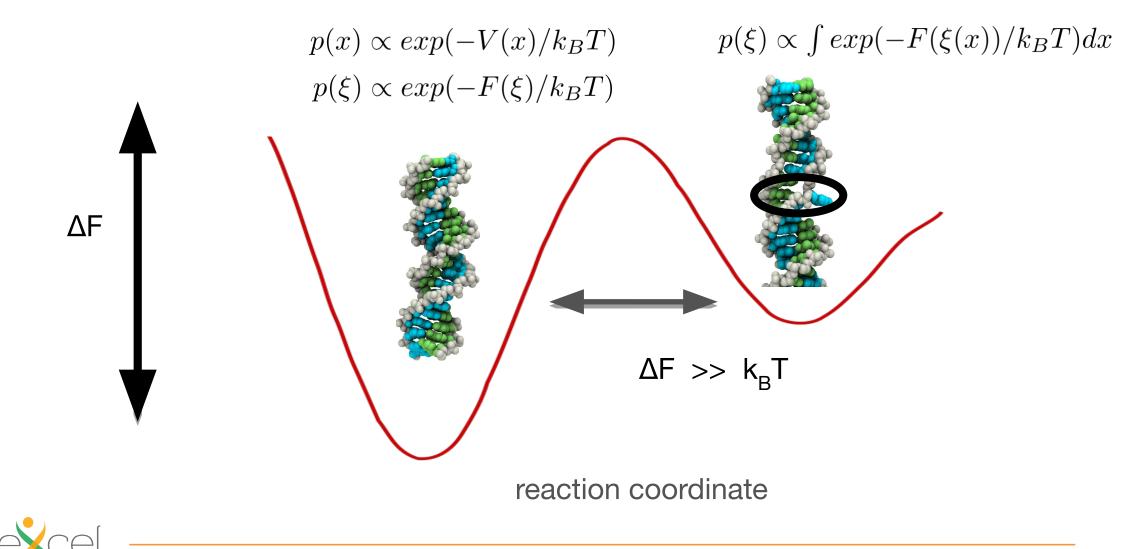
 $p(x) \propto exp(-V(x)/k_BT)$



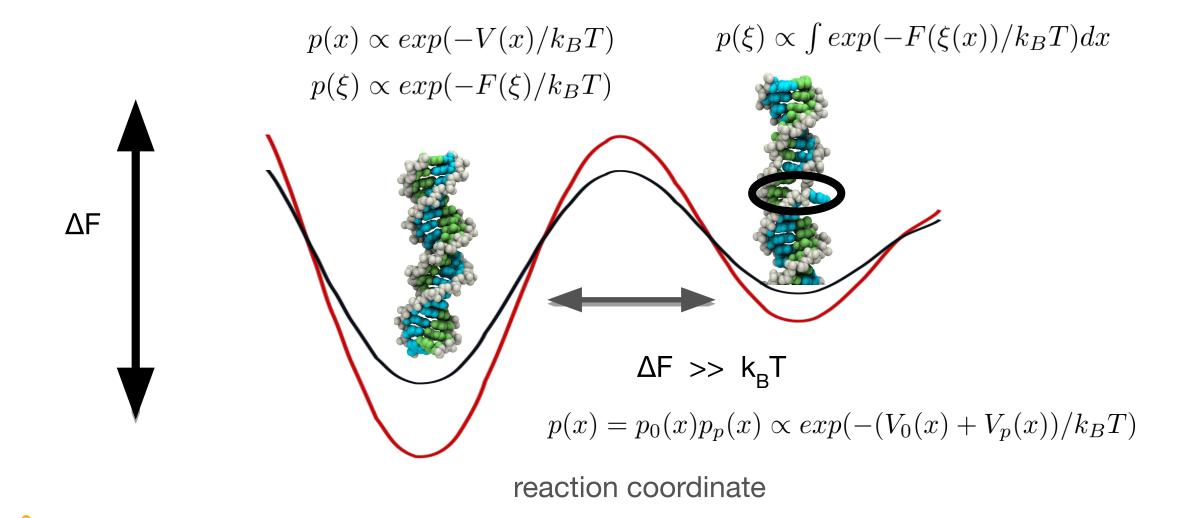
Sampling a Boltzmann distribution:



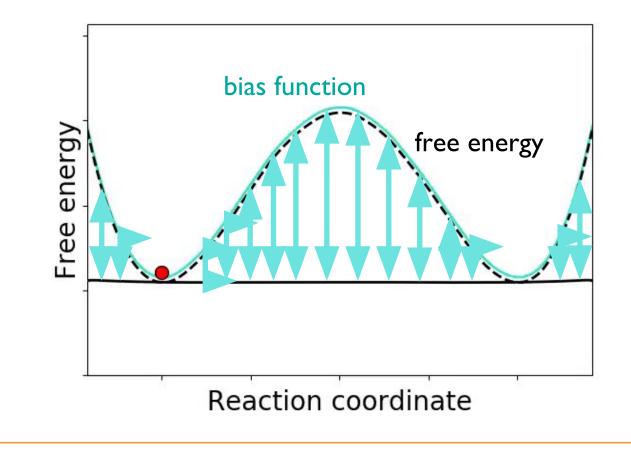
Sampling a Boltzmann distribution:



Sampling from a different Boltzmann distribution

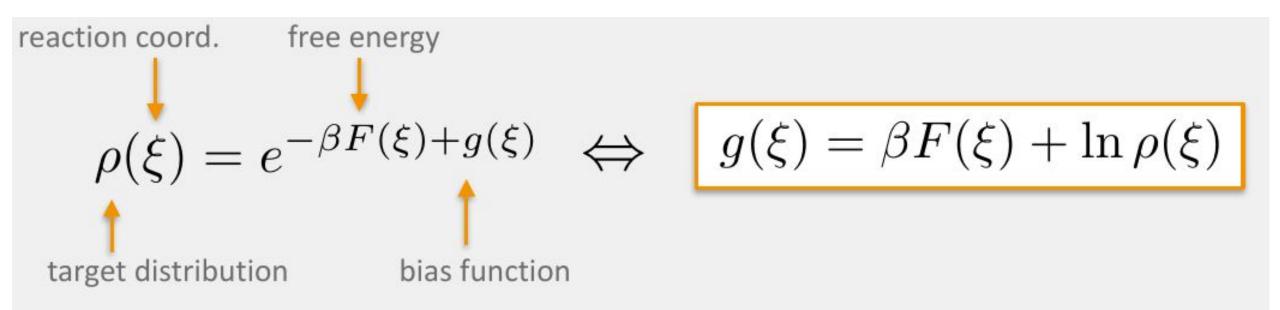


Trick: add a bias potential to make the effective potential flat Issue: the potential (or free-energy) is what we are after!





Need the free energy to apply the right bias

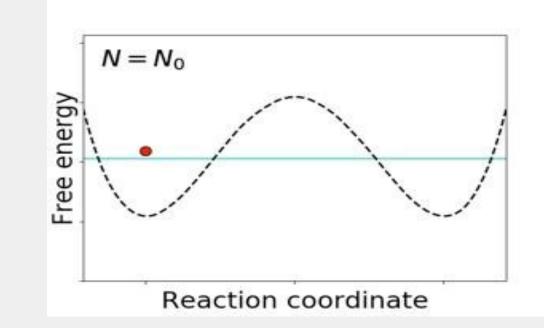


Calculating the free energy requires the bias, the bias requires the free energy — proceed adaptively!



Adaptively estimate free energy and applies the bias

- 1. Estimate free energy
- 2. Set bias
- 3. Collect (biased) samples
- 4. Update free energy estimate



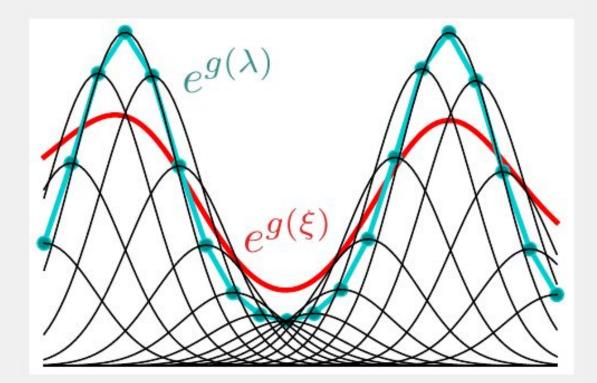


How is the bias represented and applied?

- Discretize ξ space with grid points λ
- Parameterize bias using Gaussian "basis" functions*

$$e^{g(\xi)} = \sum_{\lambda} e^{g(\lambda)} e^{-\frac{1}{2} \left(\frac{\xi - \lambda}{\sigma}\right)^2}$$

- Width σ sets resolution
- Bias force: $\nabla g(\xi(x))$



Physical interpretation of λ

$$e^{g(\xi)} = \sum_{\lambda} e^{g(\lambda)} e^{-\frac{1}{2} \left(\frac{\xi - \lambda}{\sigma}\right)^2}$$

Probabilistically the same as a having a "particle" at λ that experiences an external bias $g(\lambda)$ and interacts with ξ through a harmonic potential

$$U(\xi,\lambda) = \frac{k}{2} (\xi - \lambda)^2, \qquad k = \frac{\beta}{\sigma^2}$$

force constant
$$\lambda \text{ stays close to } \xi \text{ for large force constants:}$$
$$P(\lambda|\xi(t)) \propto e^{g(\lambda) - \frac{1}{2}\beta k(\xi(t) - \lambda)^2}$$
$$P(\lambda|\xi(t)) \quad \xi(t)$$



The algorithm again, now with more detail

"

1. Estimate free energy 2. Set bias 3. Collect samples 4. Update free energy

estimate
$$F_n(\lambda) \approx F(\lambda)$$
 exact $e^{-\beta F(\lambda)} = \int e^{-\beta F(\xi)} e^{-\frac{1}{2}(\frac{\xi-\lambda}{\sigma})^2}$ convolved free energy"

• The PMF $F(\xi)$ is extracted by an on the fly reweighting procedure

The algorithm again, now with more detail

1. Estimate free energy 2. Set bias 3. Collect samples 4. Update free energy

$$g_n(\lambda) = \beta F_n(\lambda) + \ln \rho(\lambda)$$

bias free energy estimate target



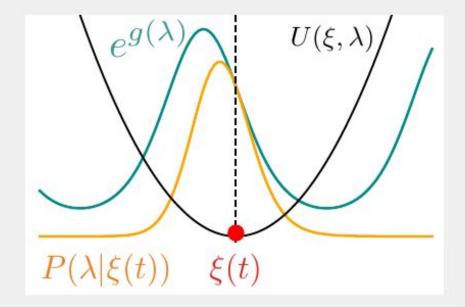
The algorithm again, now with more detail

1. Estimate free energy 2. Set bias 3. Collect samples 4. Update free energy

sample at time t, bias gn

$$w_n(\lambda|\xi(t)) = P_n(\lambda|\xi(t))$$
$$= e^{g_n(\lambda) - \frac{1}{2}\beta k(\xi(t) - \lambda)^2}$$

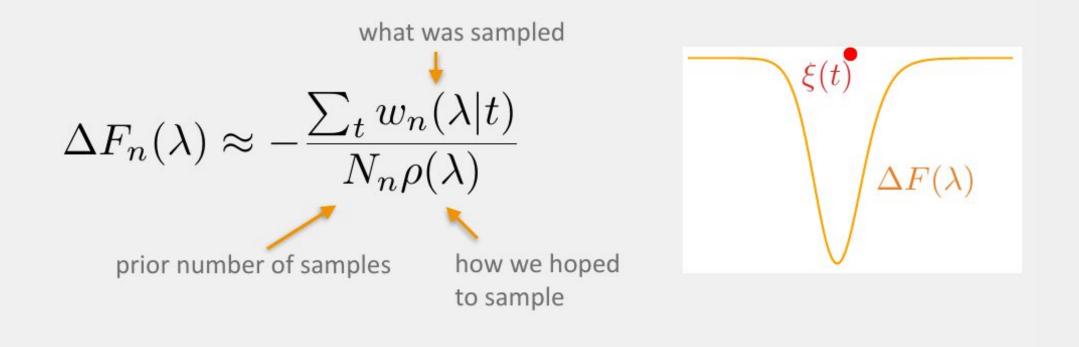
a "biased" Gaussian





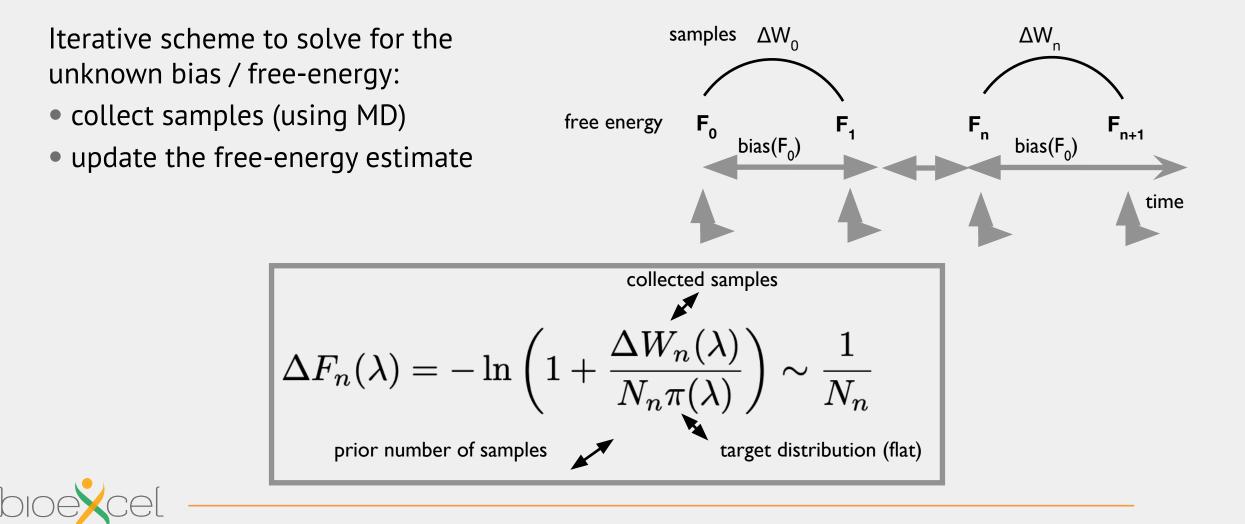
The algorithm again, now with more detail

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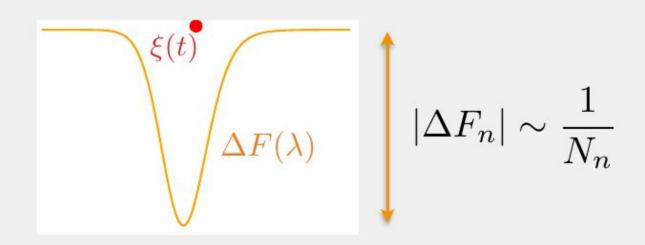




Accelerated Weight Histogram Method



The free energy update size



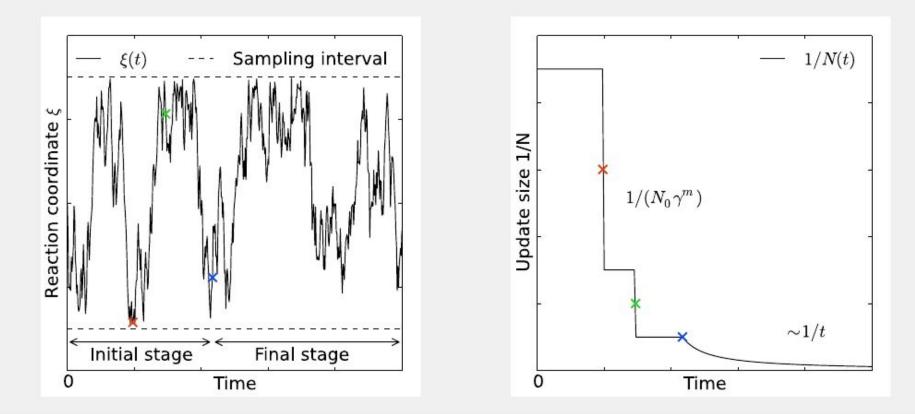
- N ~ total number of prior samples
- · reflects the accuracy of the free energy estimate
- should grow at sampling rate

$$N_n \sim t$$



Adding robustness — the initial stage

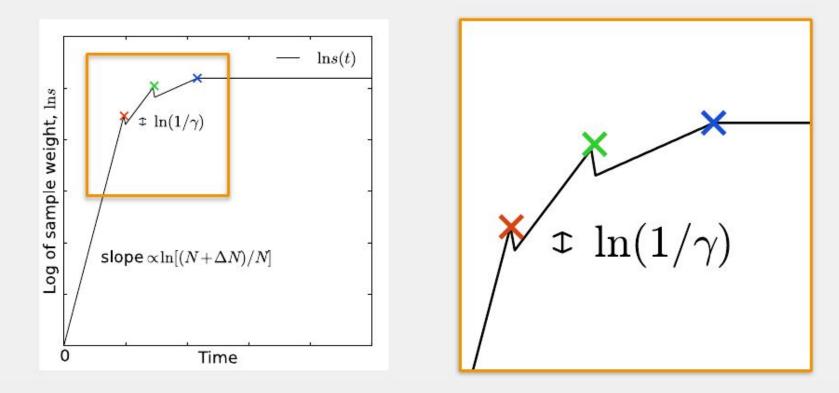
- · Letting N grow at "naturally" decreases the update size too rapidly initially
- The initial stage: keep the update size large for the first few transitions.
- After each covering of the sampling interval $N_{n+1} = \gamma N_n$



bioexcel

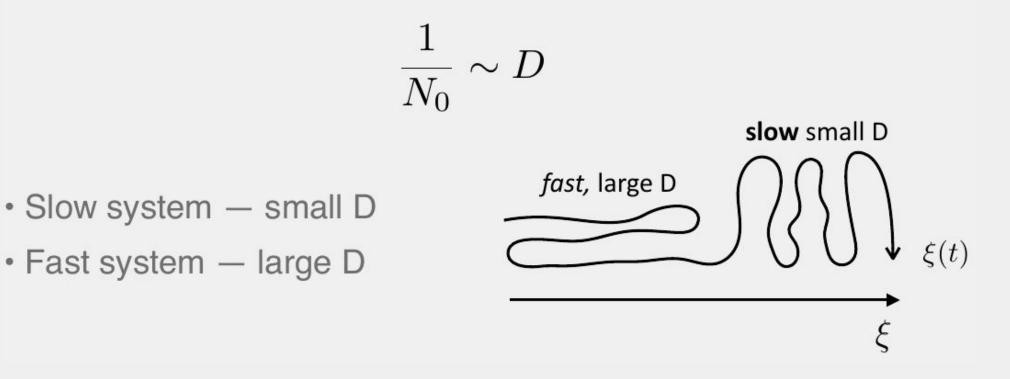
Exiting the initial stage

- Scaling N is corresponds to rescaling the current sample weight
- Scaling up, $N_{n+1}=\gamma N_n$, decreases sample weight
- Scaling down, $N_n = \text{const.}$, *increases* sample weight
- · Exit initial stage when sample weight is no longer increasing



The initial update size $|\Delta F| \sim \frac{1}{N}$

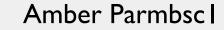
- · Sets fluctuation of the free energy and the bias.
- Slow regions get larger fluctuations than fast ones (given N)
- Parameterized in AWH by a diffusion constant D

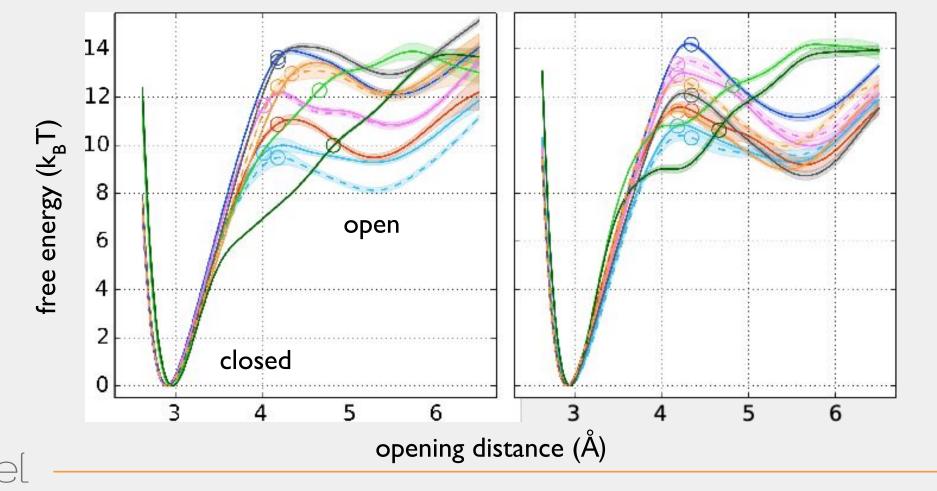




Results for different force fields (models) and base pairs

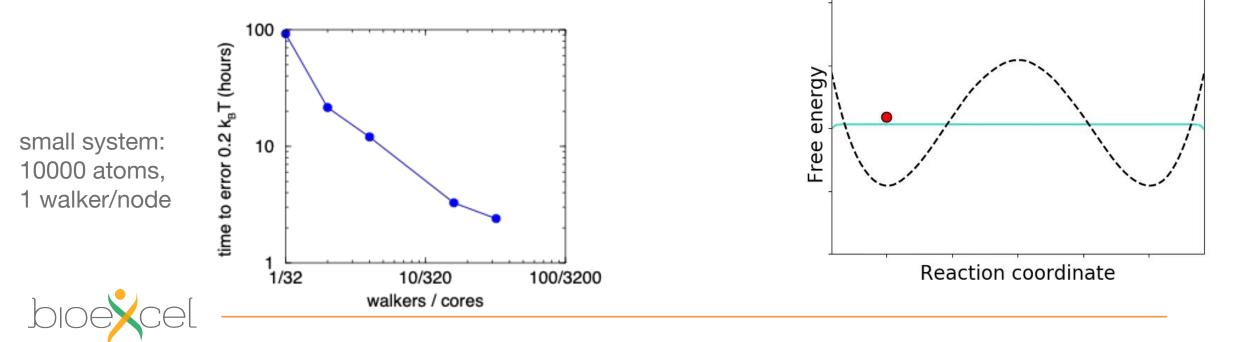
CHARMM



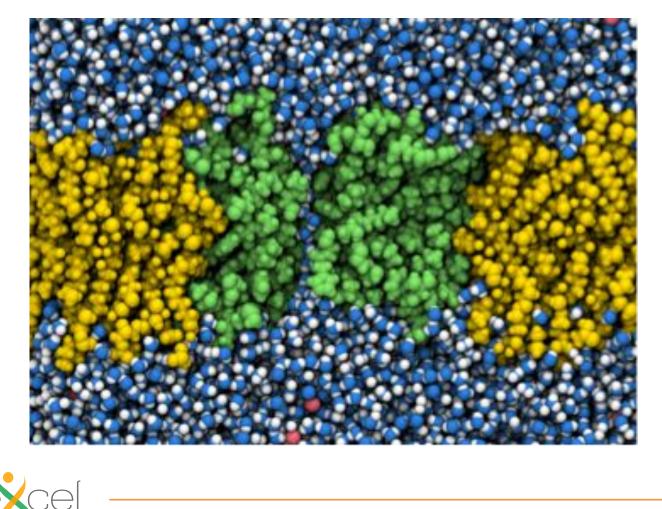


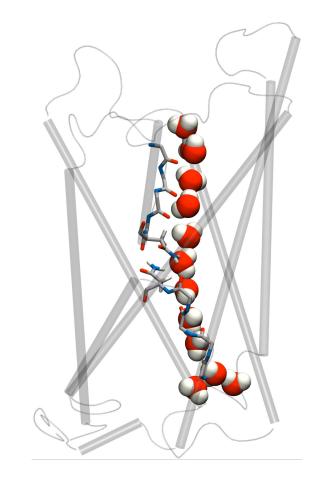
AWH with ensemble parallelism

- AWH can give exponential acceleration of barrier crossings
- We can also use multiple walkers:
 - typically exchange data every 100 steps
 - note: at 0.5 ms per step this is every 50 ms



Membrane protein: Aquaporin selectivity for water vs ammonia





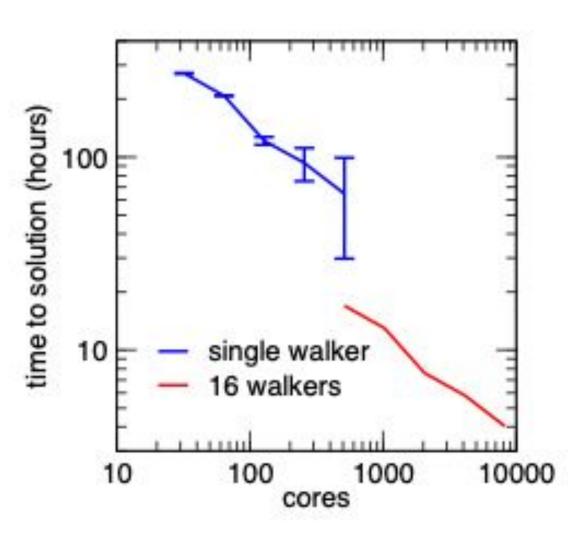


Aquaporin performance

system size: 80000 atoms

Using 16 walkers instead of 1:

- 4 times faster at 512 cores (avoids bad strong scaling)
- Time to solution at 8192 cores down to 4 hours!





BioExcel Partners 2019





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